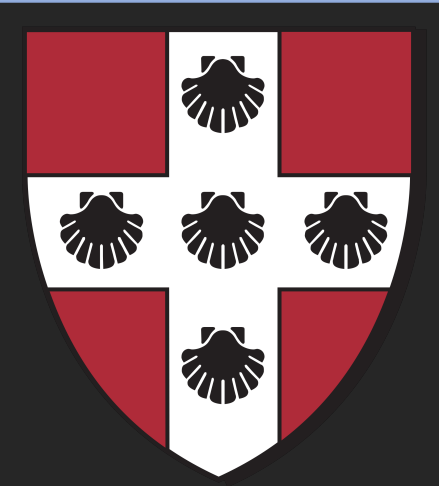




Investigating the Stability of LigAB's Dimer Interface by Molecular Dynamics Simulations

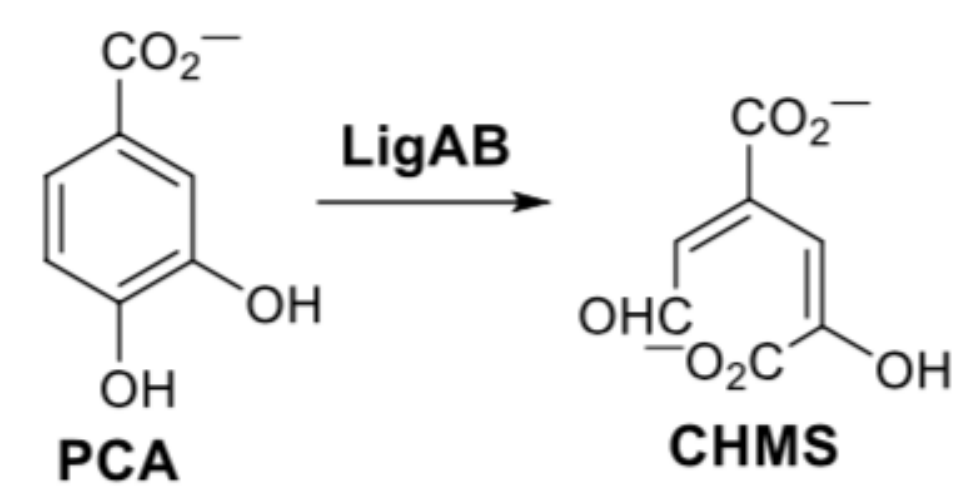
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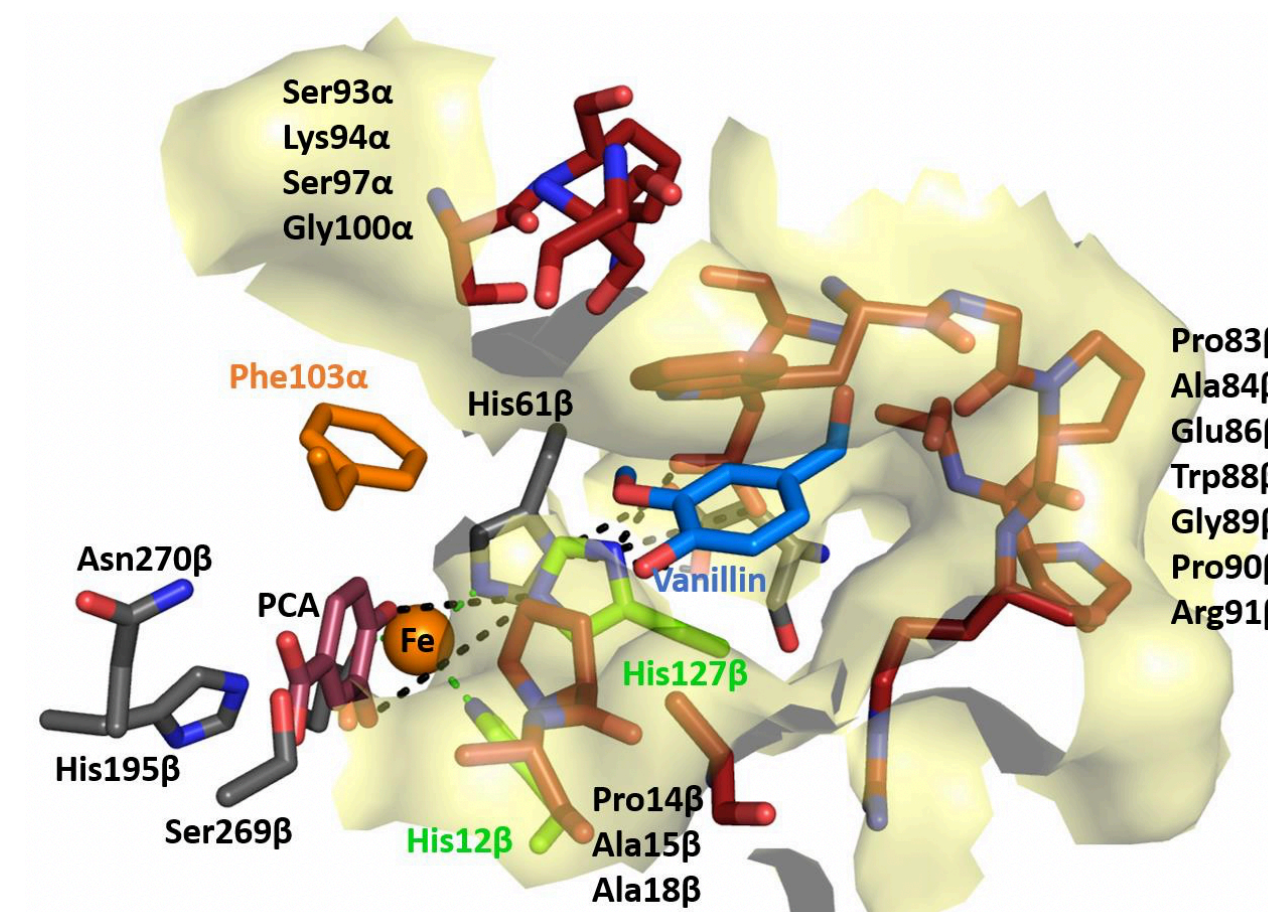


Introduction

- Lignin, a carbon compound in cell walls, is underutilized
- LigAB catalyzes ring cleaving in lignin degradation pathway



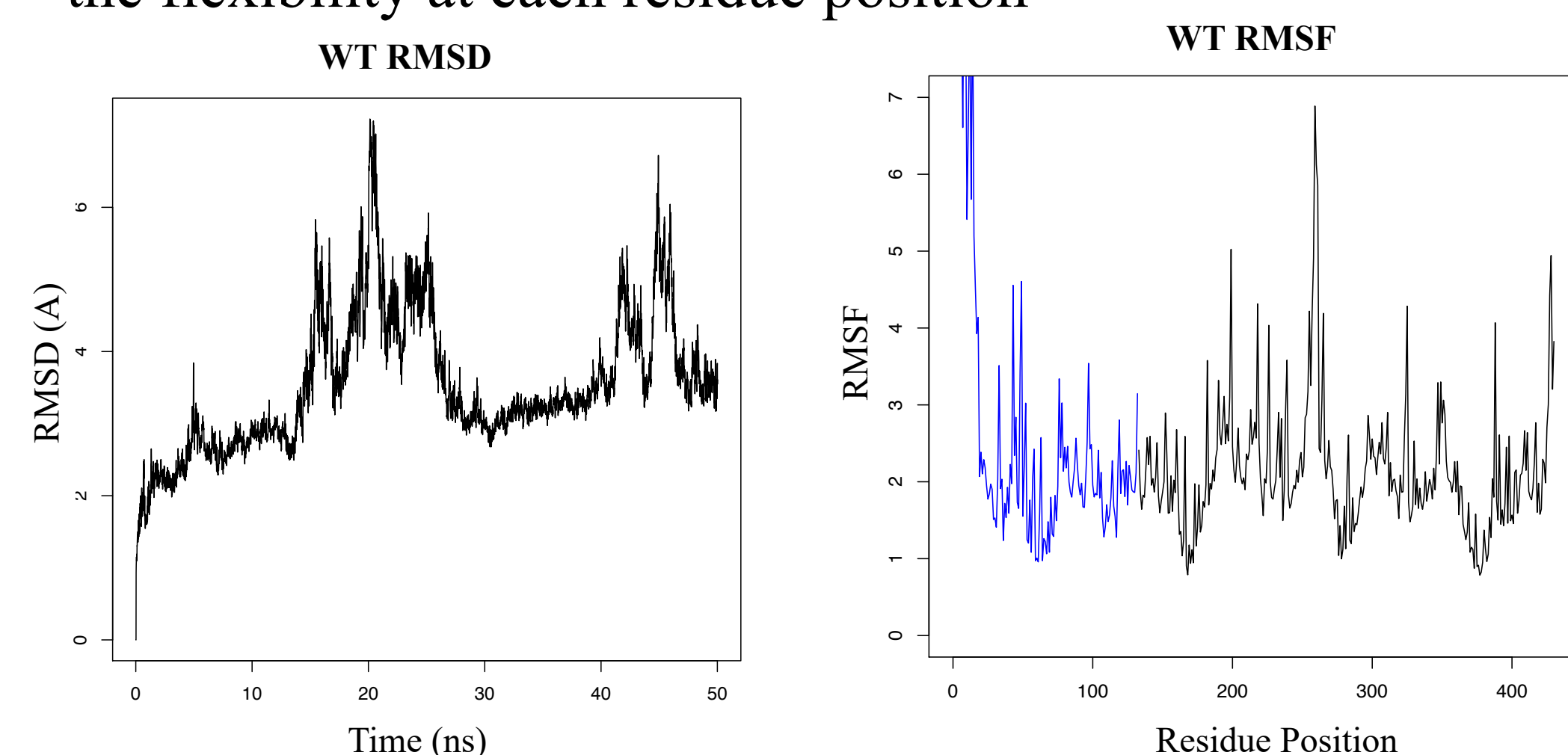
- LigAB is a homodimer of heterodimers
 - Both allosteric pocket and active site are proximal to dimer interface; dimer stability likely influences enzyme activity
 - Phe103 α residue common to both of these sites



- Previously, some Phe103 α mutants did not co-purify
 - Absence of a large, nonpolar residue disrupts interface
- Using GROMACS molecular dynamics package, stability of several Phe103 α mutants computationally determined
- Calculated binding free energy of mutants with AMBER

Wild Type

- Generated 50 ns trajectory for WT LigAB
 - Explicit solvent system and neutralized with Na⁺
- Root Mean Squared Deviation (RMSD) shows deviation of the model from starting position over time
 - Relatively small values indicated accurate model
- Root Mean Squared Fluctuation (RMSF) used to determine the flexibility at each residue position



Acknowledgements

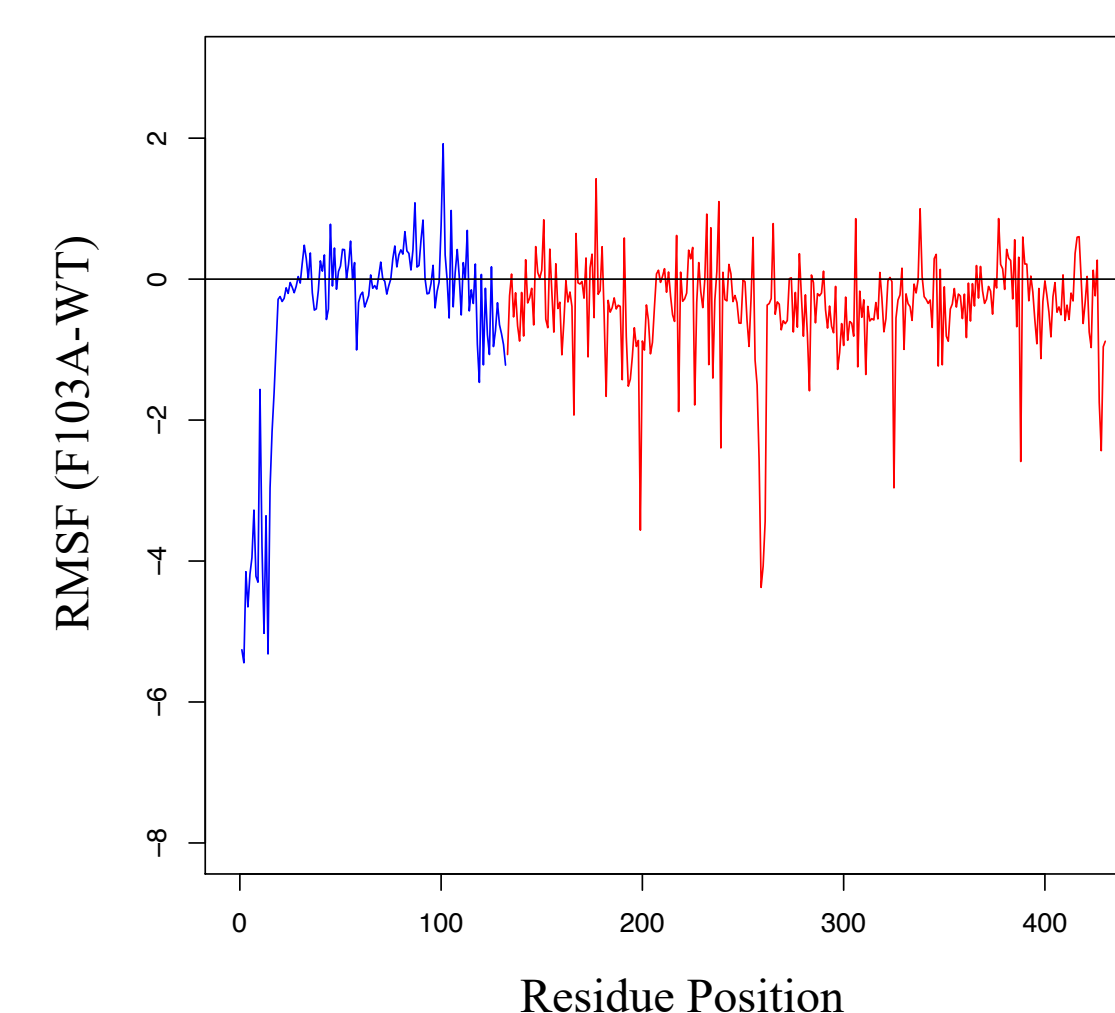
I would like to thank Bakar Hassan for guiding me through what was a more arduous journey than I ever expected. I would also like to thank Jozafina Milicaj and Angelika Rafalowski for their help trying to find old data files that might not even exist. Lastly, of course I must thank Professor Taylor for her unending support both within and outside of the lab.

F103A Mutant

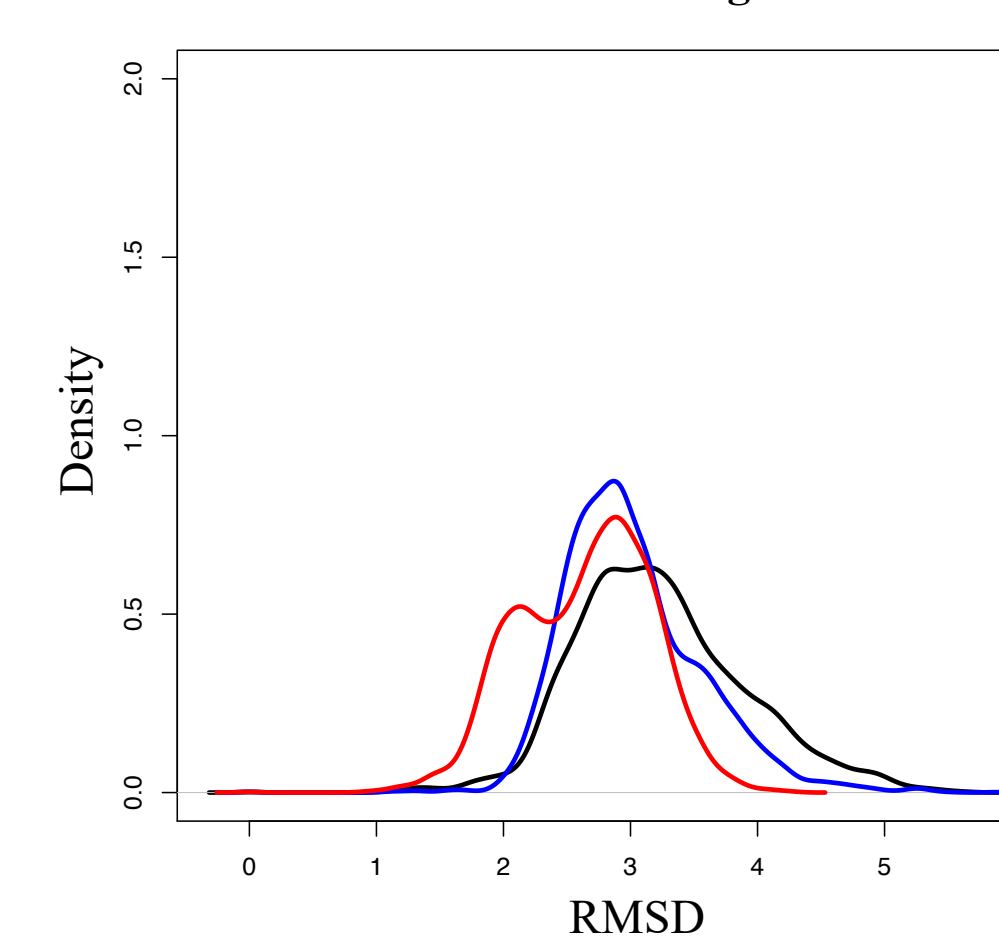
- F103A previously observed to not co-purify in expression screen
- RMSD histogram shows less deviation in beta subunit than alpha
 - Similar patterns to WT
 - Additional peak at lower RMSD for beta subunit
- RMSF of F103A smaller in beta subunit
 - Alpha mutation has global effects
- More stable than WT at residues in active site and allosteric pocket



RMSD Difference Between F103A and WT



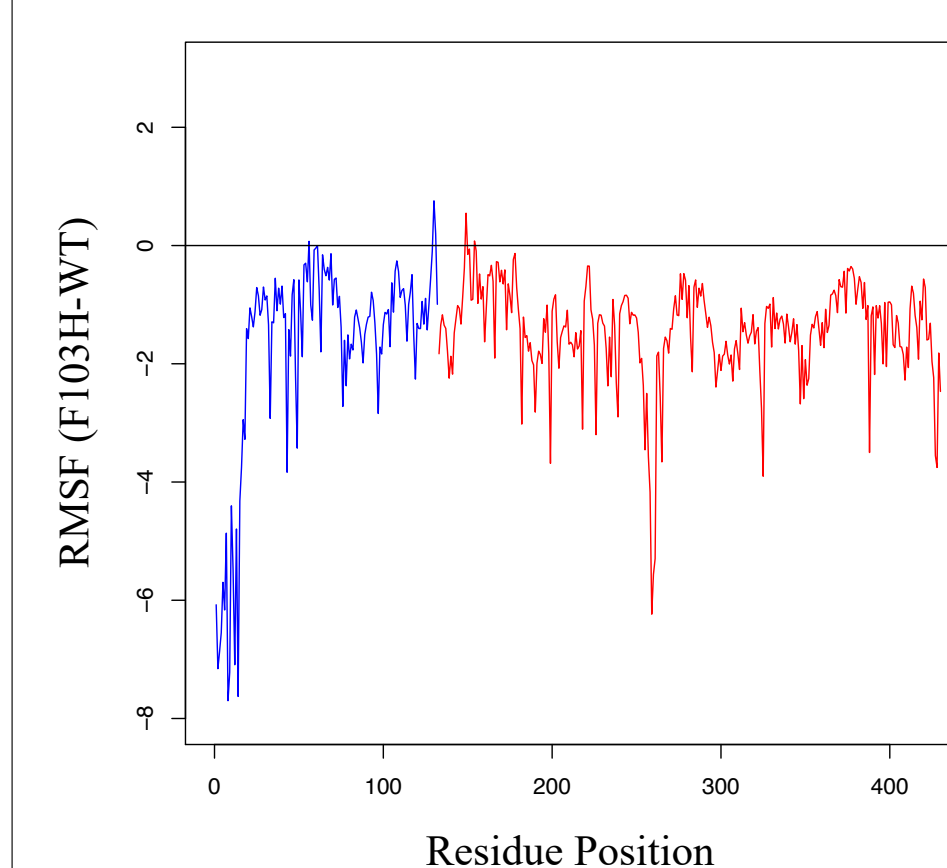
F103A RMSD Histogram



- Mutation also caused changes in cross correlational interactions across the entire enzyme (data not shown)
- These data must be analyzed with experimental data to determine which interactions might be most important for maintaining dimer stability

F103H Mutant

RMSD Difference Between F103H and WT



Active Site Residue	RMSF Difference (Mutant - WT)	
	F103H	F103A
His12	-1.24	-0.34
His61	-1.79	-0.81
His127	-4.17	-2.57
His195	-1.32	-0.55
Leu197	-1.06	-0.22
Glu242	-0.44	-0.17
Ser269	-0.95	-0.03
Asn270	-1.00	-0.23
Thr271	-1.68	-0.47

- F103H showed similar patterns to F103A
 - Less flexible in beta subunit at comparable residue positions
 - Many peaks seem to correspond to active site residues or are within proximity of the site (data for both mutants displayed in table)

References

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Free Energy Calculations

- Free energy calculations performed using MMPBSA function
 - Approximated dimer as receptor-ligand complex
- Binding free energy calculated by combining gas phase contributions with solvation free energy
- Used an implicit solvent system; Poisson Boltzmann (PB)
 - Represent solvent as continuous medium
 - Describes electrostatic environment in solvent with ions

$$\vec{\nabla} \cdot \left[\epsilon(\vec{r}) \vec{\nabla} \Psi(\vec{r}) \right] = -4\pi\rho^f(\vec{r}) - 4\pi \sum_i c_i^\infty z_i q \lambda(\vec{r}) e^{-\frac{z_i q \Psi(\vec{r})}{kT}}$$

Energy Component	WT		F103A	
	10-30ns	30-50ns	10-30ns	30-50ns
Van der Waals	-211.01	-221.05	-195.38	-195.67
Electrostatic	-556.66	-608.08	-407.44	-473.36
Electrostatic - Solvation (PB)	634.34	687.19	493.87	564.51
Non Polar Cavity Formation	-147.59	-155.41	-134.42	-137.91
Non Polar Dispersion	286.39	296.42	262.05	267.45
Total Binding Free Energy	5.465	-0.936	18.676	26.023

- Total binding free energy of F103A is significantly less stable than WT; not within range of standard deviations
 - Confirms original hypothesis
- Electrostatic interactions overall weaker in F103A
 - Both attractive and repulsive forces
 - Might implicate water in stability of the dimer interface or disruption of the stabilizing interactions

Future Directions

- Trajectories of at least 50 ns will be generated for all previously studied Phe103 α mutants
- Experimental data will be collected for each mutant
 - Aerobic purification and corresponding SDS-PAGE gels
 - Example below shows F103S; beta subunit (32 kDa) elutes without the alpha subunit (18 kDa)
 - Resulting gels will be quantified to determine co-purification of respective mutants
- Gives insight on which interactions prevent dimer formation

